

BIOGAS PRODUCTION FROM ANIMAL AND MODIFIED OIL PALM BUNCH WASTES



*Ayedun Hassan¹, Adeyemo Isaiah Adeyemi², Ayadi Peter Odunayo¹

- 1. Department of Chemical Sciences, Olusegun Agagu University of Science and Technology, P. M. B 353, Okitipupa, Ondo State, Nigeria.
- ². Department of Biological Sciences, Olusegun Agagu University of Science and Technology, P. M. B 353, Okitipupa, Ondo State, Nigeria.

 $*corresponding\ Author\ E-Mail: \underline{ht.ayedun@oaustech.edu.ng}$

Received: May 18, 2023 Accepted: July 10, 2023

Abstract

Increase in prices of cooking gas and the need to keep our environments clean makes research on biogas from wastes very useful. The study was carried out to use animal wastes mixed with empty bunches of oil palm fruits (EBPF) to generate gases that can be used for cooking. The cow dung was mixed with EBPF in the ratio 4:0, 4:1, 2:1 and 1:1. The microorganisms were isolated using standard method and allowed to act on it over 5 weeks at room temperature, 27°C, 30 °C and 35 °C. The gases collected were subjected to laboratory analysis using Gas Chromatography (GC) with a thermal conductivity detector. The results showed that average % by volume of methane recorded for each of the ratio are 57.6 \pm 1.74 %, 52.2 \pm 1.56 %, 25.3 \pm 2.38 % and 9.66 \pm 0. 817 % respectively. The more EBPF added the less gas of interest produced. Analysis of residue showed % N, P, K, and Mg to be 0.409 \pm 0.349 %, 0.113 \pm 0.126 %, 0.978 \pm 0.817 %, and 0.224 \pm 0.192 % respectively which implies materials that can be incorporated in soil amendment. Removing empty bunches of oil palm fruits from the environment to generate gas reduces environmental pollution caused by burning.

Keywords:

Biogas, Cow wastes, Cellulose, Delignification, Oil palm waste, Residue

Introduction

The quest for the reduction of greenhouse gasses and the need for sustainable energy propelled the research interest into alternative fuels. The world is embracing renewable energy due to the depletion of fossil fuel and the adverse effect on of its use on the environment (Haider, 2020). The menace of environmental pollution and climate change has been associated with the use of fossil fuels. The developed nations have been using renewable energy resources while the developing Countries are still lacking behind (Donkin et al., 2013). Nigeria has abundance of animals and birds that produced wastes which are thrown into open space, causing environmental pollution and health issues in the communities. These wastes, if properly harnessed, can be source of energy for urban and rural inhabitants (Samani et al., 2017). Production of biogas from cow dung is preferred because energy derived from it is environmentally friendly (Putri et al., 2012).

Cellulosic material has wide application, such as biofuels, macromolecules, polymeric and monomeric aromatic compounds provided the lignin components is removed or extracted (Iskalieva *et al.*, 2012; Varanasi *et al.*, 2013;). Wood and other agricultural residues consists of cellulose, hemicelluloses and lignin (Zhang et al., 2019). Little attention was given to lignin as a compound due to its complexity and difficulties in chemical modification, however, emergence of its application is found in cosmetic, pharmaceutical, and food industries.

Biogas produced through anaerobic digestion (AD) is environmentally friendly and energy efficient, when compared to other forms of energy (Achinas *et al.*, 2017). Anaerobic digestion is more preferred because of low cost and its efficiency (Nizami and Murphy, 2010).

Generally, all kinds of biomass can be used as substrates for the biogas production, however, appropriate pretreatment, digester condition is necessary for effective biogas production (Hagos *et al.*, 2017). Substrates such as animal wastes, sludge, waste waters, lignocellulosic materials, crop residue and grass silage were used (Moller *et al.*, 2004; Bohn *et al.*, 2007). Rumen microbes (bacteria, fungi, protozoa) are a valuable protein source and can supply 60 to 70 per cent of the animal's protein requirement. Microbiota present in the rumen of cows allowed them to digest fibers, synthesize vtamins, converts non-proteins nitrogen into protein and digest toxic substances. (Krause *et al.*, 2013).

Different researchers have also reported various species of fungi in cow dungs. Asao et al. (1993), reported the presence of different fungi species in the gut of ruminants and confrmed that anaerobic fungi and enzymes play a vital role in the degration and assimilation of fibrous feeds consumed by animals. Brownlee et al. (1996), confirm the presence of Trichoderma reesei, Aspergillus niger and Penicillium italicum, in the gut of cows using DNA and PCR to probe the fungal population while several researchers also reported the presence of fungi including Phanerochaete chrysosporium, Rhizopus stolonifera, Mucor mucedo and Fusarium oxysporum in the guts of ruminants and invariably their dungs (Azad et al., 2020). The ability of these fungi to digest cellulose in the grasses being fed upon by cows makes them to be in a good symbiotic relationship with the guts of

Most of the biogas researchers in Nigeria used animal dung as substrate, (Ekka *et al.* 2016). A few have explored other substrates such as ornamental plant (Garba, 2002), cassava peels (Adelekan and Bamgboye, 2009), Onion bulbs, and Protein-rich biomass (pig blood) (Kovacs *et al.*, 2013). Some studies have been carried out on conversion of palm oil mill

wastes to useful material (Adela et al., 2014) but few focus on biogas generation especially the use of empty bunches of oil palm fruits.

Okitipupa is an agricultural based Local government where biomass wastes generation from oil palm processing, plantain peels, cassava peels and other crop residues are generated in large quantities. Animal waste such as cow dung, poultry droppings, and pig waste were also generated in substantial amount and disposed on land surfaces. All these can be harnessed into a useful source of renewable energy. The burning of empty bunches and other solid wastes constitute air pollution. There is need to find a way of converting these wastes to a valuable resource. The objective of the study is to convert empty bunches of oil palm fruits to biogas thereby circumventing burning which is the usual practice in the area.

Materials and Methods

Materials used include fresh cow dung and empty palm fruit bunches.

Waste Collection: Fresh cow dung are collected from Agric Reality Farm at Maclean village off Aye road, Okitipupa, Ondo State, Nigeria. The empty palm fruit stalk bunches were collected from an Oil mill factory located in Ayeka, Igbodigo, Okitipupa, Ondo State. The cow dungs were collected using sterile polythene bags immediately they are excreted, labelled, and taken to the laboratory for analysis.

Bacteria and Microbial screening: Spread plate technique was used for the isolation and different culture characteristics were observed after incubation. Total plate count of bacteria in each sample was determined using a colony counter and calculated as colony forming units per ml (CFU/ml) with the formula:

CFU/ml = No. of colonies x Dilution factor/ volume of inoculum.

Streak plate method was used and the isolated colonies were individually sub cultured to obtain pure culture. Differential tests and biochemical test was carried out. Biochemical test used to identify the isolates are catalase test, citrate test and urease test. Isolated fungal colonies were identified for both microscopic and macroscopic examinations (McGinnins and Borgers, 1980).

Pretreatment of empty oil palm bunches (EBPF): The empty bunches of oil palm fruits were dried and milled to reduce the particle size. Delignification was done by boiling the oil palm wastes in 1.5M NaOH for 2 hours. It was cooled to room temperature, filtered washed with IL of hot distilled water. The solid were dried in an oven for 48 hours at 45 °C milled into powder. Delignification by alkaline pretreatment with 2 - 4% NaOH in autoclave at 121 °C has been suggested (Pramasari *et al.*, 2021; Lourenço *et al.*, 2021).

Slurry preparation: The modified method used by Ahamed et al. (2016), was adopted for calculation. The slurry of total weight 2 kg was prepared by weighing 500g of cow wastes and 140g of delignify empty bunches of oil palm fruits (EBPF) while 1360g of water was added. The cow wastes, EBF and water represented 25 %, 7% and 68% of the total

slurry respectively to achieve 4: 1 of waste to EBF ratio. The weight was adjusted to obtain other ratio used for the experiment.

Digestion and biogas generation: The batch digester was set while the connection were tightly fitted to prevent air leakage. The material remained in the digester throughout the entire digestion period, no new fresh substrate was added and no digest residue was removed during the process. The methane yield was calculated from % volume recorded. The assumption adopted was that, at standard temperature and pressure, 1g of oxygen demand take 400mL (0.4 m³ required 1kg of oxygen demand) of methane (Hamilton, 2022).

Analysis of the gas and the residues:

Gas Chromatography – Mass Spectrometry (GC-MS) (Perkin Elmer) was used to determine different components of gases produced with a thermal conductivity detector (TCD). Atomic Absorption Spectroscopy (AAS) (Bulk Scientific model) was used for determination of metals in the residue. Heavy metal contents (Cd, Mn, Ni, Zn, Cu, Fe) in the biogas residues were determined using Atomic Absorption Spectroscopy (AAS) (APHA, 1998).

Results and Discussion

The results of bacterial loads on cow dungs are presented (Table1). Gut of ruminants have been widely reported to accommodate various microflora because the foregut of ruminants houses an ecosystem of micro-organisms that breakdown plant cell wall. It allows the animal to obtain nutrients from both the plant material and the microbes themselves (Abecia *et al.*, 2013). The gram positive bacteria are stained dark blue, while gram negative bacteria appeared pink red.

Table 1: Microbial Loads on cow dungs

S/N	Stock	10-3	10-5
1	175×10^7	12.0	3.0
2	167 x 10 ⁷	12.0	3.0
3	205×10^7	14.0	3.0
4	190×10^7	13.0	4.0
5	165 x 10 ⁷	12.0	4.0
6	180×10^7	13.0	4.0
7	155 x 10 ⁷	12.0	3.0
8	160 x 10 ⁷	12.0	3.0
9	105×10^7	11.0	3.0
10	165×10^7	13.0	4.0

Bacillus cereus, Escherichia coli, Clostridum perfringes, Pseudomonas aeruginosa Salmonella sp, Staphylococcus aureus, Streptomyces thermos-autotrophicus, and Alcaligenes faecalis were identified and isolated from the cow dung used in the present study (Table 2). This is in agreement with previous studies (Hawkes et al., 2008). Different species of Bacillus such as Escherichia coli, Proteus mirabilis, Streptomyces spp, Pseudomonas sp, Salmonella sp, Staphylococcus aureus, and Alcaligenes faecalis have been isolated from cow dungs (Kartikey et al., 2015; Adeyemo and Waleola 2016; Cole et al., 2022).

Table 2: Results of Biochemical tests on bacterial isolates from cow dungs

Tubic 24 Itesuits of Biochemical tests on caeterial isolates from 40 % dailigs											
Isolate	Gram	Form	Cat	Cit	Ure	Oxi	Lac	Glu	Suc	Mal	Suspected Organism
	test										
1	+	Rod	+	+	+	+	+	+	Variable	+	Bacillus cereus
2	-	Rod	-	-	-	-	+	+	Variable	+	Escherichia coli
3	+	Rod	-	+	-	-	+	+	+	+	Clostridum
											perfringes
4	-	Rod	+	+	-	+	-	-	-	-	Pseudomonas
											aeruginosa
5	-	Rod	+	+	-	-	-	+	-	-	Salmonella sp
6	+	Cocci	+	+	-	+	+	+	-	+	Staphylococcus
											aureus
7	+	Filament	+	+	+	+	+	+	+	+	Streptomyces
											thermoautotrophicus
8	-	Rod	+	-	-	+	+	+	+	-	Alcaligenes faecalis
9	-	Rod	+	+	+	-	-	-	-	-	Proteus mirabilis

Key: Gram test – Gram staining test, Form- shape under microscopic view, Cat – Catalase test, Cit – Citrate test, Ure- Urease test, Oxi- Oxidase, Lac- Lactose, Glu- Glucose, Suc- Sucrose, Mal- Maltose, + = positive, - = negative, -/+ = variable.

The Clostridium genus is involved in hydrolysis of the substrates, acidogenesis and acetogenesis stages of anaerobic digestion (Tapadia - Maheshwari *et al.*, 2019). *Escherichia coli* were found in lactose and glucose and maltose but variable in sucrose. The few presence of E. coli is due to killing effect of anaerobic digestion. This is consistent with findings of Ye *et al.* (2012). Furthermore, increase in temperature can easily destroy E. Coli and caused its rapid death (Liang *et al.*, 2021).

With the exception of the first preparation without EPFB, other slurry was prepared such that the total weight of the slurry was 2 kg. With the assumption of Hamilton, (2022), that 1 kg of oxygen demand (OD) take 400 mL of methane and 1kg OD remove $0.4 \, \mathrm{m}^3$ of methane produced. Fifty percent (50 %) cow dung with no EPFB and 50 % of water yielded $2.86 \times 10^{-3} \, \mathrm{m}^3$ of methane (Table 3). Twenty-five percent (25 %) cow dung with 7 % EPFB and 68 % of water yielded $2.61 \times 10^{-3} \, \mathrm{m}^3$ of methane. Twenty-five percent (25 %) cow dung with 12.5 % EPFB and 62.5 % of water yielded $1.26 \times 10^{-3} \, \mathrm{m}^3$ of methane. Twenty-five percent (25 %) cow dung with 25 % EPFB and 50 % of water yielded $0.483 \times 10^{-3} \, \mathrm{m}^3$ of methane.

Table 3: mass of slurry with quantity of materials used

		Cow		Methane
Cowdung		dung:	Water	yield m ³
(g)	EPBF (g)	EPBF	(g)	(x10-3)
500	0	4:0	500	2.86
500	140	4:1	1360	2.61
500	250	2:1	1350	1.26
500	500	1:1	1000	0.483

Significantly high percentage of methane (57.6 ± 1.74) was recorded when 100% of cow dung was used (4:0) (Table 4). The composition of biogas and the intensity of gas formation are heavily depending on the quantity of the animal wastes, viability of micro-organisms present in the waste and temperature (Odonkor and Mahami, 2020).

Table 4: Results of gas produced at different ratio of cow wastes with EBPF

Cow					
Dung	CH ₄	NH_3	CO	H_2S	CO_2
:	(%	(%	(%	(%	(%
EBPF	volume)	volume)	volume)	volume)	volume)
	57.6	0.209	0.718	0.796	30.85
4:0	$\pm 1.74c$	$\pm 0.045b$	$\pm 0.075d$	±0.036c	±1.20d
	52.2	0.048	0.439	0.409	21.4
4:1	±1.56c	$\pm 0.038a$	$\pm 0.0376c$	$\pm 0.052b$	$\pm 0.832c$
	25.3	0.015	0.241	0.169	8.55
	$\pm 2.38b$	$\pm 0.006a$	±0.022b	±0.031a	±0.42c
2:1					b
	9.66	0.016	0.047	0.106	0.387
1:1	±0.817a	$\pm 0.007a$	±0.01a	$\pm 0.0147a$	±0.22a

Note: Data with the same letter down the column are not significantly different using Duncan Multiple Range Test.

There is no significant difference in % methane produced when 4:1 of cow wastes to empty bunches of oil palm wastes was used and 100 % of cow wastes (4:0) was used at the temperature of 33°C and pH range of 6.8 to 7.5. This agrees with pattern of results reported when sheep manure was used without modification (Yoshida et al., 2020). The % volume of gases produced decreases as the quantity of cow dung reduces which agrees with the report of Putri et al. (2012). The pH of the slurry was varied between 5 and 7.5, it was found that higher gas pressure was recorded at the lower pH (Figure 1). This is consistence with findings of Jacob et al., (2018). The activity of methanogenic bacteria is more favoured by acidic condition, which also depends on when the animal was fed last (Madigan et al., 2011). Alkaline pH is toxic to the bacteria which decreases the activity due to conversion of ammonia to ammonium, hence lower gas pressure was recorded (Chen et al., 2008).

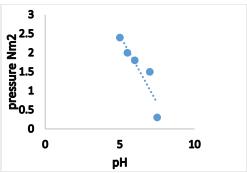


Figure1: Variation of pH with gas pressure.

In this experiment, three temperatures 27°C, 30°C and 35°C were used corresponding to the laboratory environment, under the shed constructed and outside the shed. The gas pressure recorded increases with increase in temperature (Figure 2). At 27°C, the gas pressure of 1.05Nm² was measured, at 30°C, the gas pressure of 1.6 Nm² was recorded while at 35°C, the pressure of 2.05Nm² was recorded. Most methanogenic bacteria are mesophilic and thrive in conditions that resemble their original habitat which is cow rumen

(Mussoline et al., 2013).

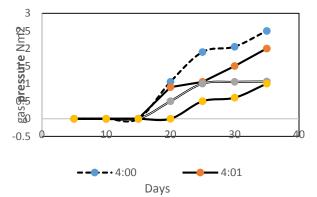


Figure 2: A graph of pressure monitored at different days for different preparations

No visible changes was recorded in the pressure gauge until 20th day after incubation. With 100 % cow-dung (4:0), the pressure recorded are 1.05 Nm², 1.9 Nm², 2.05 Nm², and 2.5 Nm² for day 20, 25, 30 and 35 respectively (Figure 2). When cow-dung with EBPF in the ratio 4: 1 was used, the pressure recorded are 0.9 Nm², 1.05 Nm², 1.5 Nm², and 2 Nm² for the days respectively. With the cow-dung and EBPF in the ratio 2:1, the pressure recorded are 0.5 Nm², 1 Nm², 1.05 Nm², and 1.06 Nm² for the days respectively. Finally, with cowdung and EBPF in the ratio 1: 1, the pressure recorded are nil, 0.5 Nm², 0.6 Nm², and 1 Nm² for the days respectively. The average concentration of N, P, K and Mg in the residue are 0.409 ± 0.349 %, 0.113 ± 0.126 %, 0.978 ± 0.817 %, and 0.224 ± 0.192 mg/kg respectively (Table 5). Liang et al. (2021) recorded 0.059 %, 0.05% and 0.052% for N, P and K respectively for animal wastes fermentation residue.

Table 5: Elemental composition of residues of digested cow dung with EBPF

Elements	Min	Max	Mean \pm SD
N (%)	0.001	0.97	0.409 ± 0.349
P(%)	0.001	0.38	0.113 ± 0.126
K(%)	0.01	2.26	0.978 ± 0.817
Mg (mg/kg)	0.007	0.59	0.224 ± 0.192
Mn (mg/kg)	0.001	0.814	0.247±0.26
Zn(mg/kg)	0.001	0.621	0.209 ± 0.21
Fe (mg/kg)	3	129	64.4±45.7
Cu (mg/kg)	0.01	0.867	0.341±0.269
Cd (mg/kg)	0.001	0.011	0.005±0.004
Ni(mg/kg)	0.003	0.204	0.099±0.08

The appreciable percentage of N, P and K found in the residue is comparable with previous findings in literature (Panjaitan, *et al.*, 2022). High concentration of K and Mg found in cow dung is responsible for high activity of methanogens (Abu -Ashour *et al.*, 2010). This makes the residue a material that can be recommended for possible use as soil amendment.

Mn concentration ranged between 0.001 to 0.814 mg/kg with the mean value of 0.247 \pm 0.26 mg/kg. Zn concentration ranged between 0.001 to 0.621 mg/kg with the mean value of 0.209 \pm 0.21 mg/kg. Fe concentration ranged between 3 to 129 with the mean value of 64.4 \pm 45.7 mg/kg. Cu concentration ranged between 0.01 to 0.867 mg/kg with the mean value of 0.341 \pm 0.269 mg/kg. The elemental analysis of the residue showed the presence of some toxic elements such as Ni and Cd. Ni concentration ranged between 0.003 to 0.204 mg/kg with the mean value of 0.099 \pm 0.08 mg/kg (Table 3). Cd concentration ranged between 0.001 to 0.011 mg/kg with the mean value of 0.005 \pm 0.004 mg/kg. The presence of toxic elements in the residue showed that it can be used to land fill waste dump site in a professional way in order to prevents its migration into groundwater.

Conclusion

Results from the present study showed that cow dung can best be combined with EBPF in the ratio not exceeding minimal use of EBPF at acidic pH and moderate temperature. The EPBF can be used generate biogas for domestic use and avert health hazard arising from its open burning. The residue contains substantial amount of N, P, and K which suggest possible suitability for soil amendment to improve crop yield. The use of EBPF to generate biogas can remove waste from our environments and improve energy yield for sustainable development. Further study should be carried out on the use of residue for fertilizer to ascertain a particular crop it can support.

Conflict of Interest: There is no conflict of interest

Funding: This project was funded by TETFUND Nigeria through IBR 2020

Acknowledgement: The assistance rendered by Adefisayo Sarah Olorunsola, Ekundayo Paul Oluwashina and

Folorunsho Oluwaseun David during the experiment are acknowledged.

References

- Abecia L, MartínGarcía AI, MartínezG, Newbold CJ, & Ya nez RDR (2013). Nutritional intervention in early life to manipulate rumen microbial colonization and methane output by kid goats postweaning. *Journal of Animal Science* 91, 4832 – 4840
- Abu-Ashour J, Abu Qdais H, & Al-Widyan M (2010).

 Estimation of animal and olive solid wastes in
 Jordan and their potential as a supplementary energy
 source: An Overview. *Renewable & Sustainable*Energy Review 14, 2227–2231.
- Adela BN, Muzzammil N, Loh SK, & Choo YM (2014).

 Characteristics of palm oil mill effluent (pome)
 in anaerobic biogas digester. Asian Journal of
 Microbiology Biotechnology & Environmental
 Science
 16, 225 231.
- Adelekan BA, & Bamgboye AI (2009). Comparison of biogas productivity of cassava peels mixed in selected ratios with major livestock waste types. African Journal of Agricultural Research 4, 571-577.
- Adeyemo IA, & Waleola OA (2016). Prevalence and Incidence of *Escherichia coli* O157 in domestic goat dungs of Okitipupa township. *Nigerian Journal of Pure & Applied Sci*ence 29, 2803 2810.
- Achinas S, Achinas V, & Euverink GJW (2017). A technological overview of biogas production from biowaste. *Engineering* 3, 299 307.
- Ahamed JU, Raiyan MF, Hossain MDS, Rahman MM, & Salam B (2016). Production of biogas from anaerobic digestion of poultry droppings and domestic waste using catalytic effect of silica gel.
 International Journal of Automotive & Mechanical Engineering 13, 3503 3517, DOI: https://doi.org/10.15282/ijame.13.2.2016.17.0289 3503
- APHA (1998). American Public Health Association Standard methods for the examination of water and waste water, 19th Edition. APHA, AWWA, WEF. Washington DC. 4 - 67, 99 -144pp.
- Asao N, Ushida K, & Kojima Y (1993). Proteolytic activity of rumen fungi belonging to the genera *Neocallimastix* and *Piromyces*. *Letters in Applied Microbiology* 16, 247 250.
- Azad E, Fehr KH, Derakhshani R, Forster S, Acharya E, Khafipour E, & McGeough TM (2020).

 Interrelationships of Fiber-Associated Anaerobic Fungi and Bacterial Communities in the Rumen of bloated Cattle Grazing Alfalfa. *Microorganisms* 8. https://doi.org/10.3390/microorganisms8101543
- Bohn I, Bjornsson L, & Mattiasson B (2007). The energy balance in farm scale anaerobic digestion of crop residues at 11–37°C. *Process Biochemistry* 42, 57 64.
- Brownlee AG, Rintoul AJ, Gordon GLR, & Phillips MW (1996). Monitoring rumen fungal

- populations with PCR and DNA probes. *Microbiology Australia* 17, A38
- Cole SD, Swiderski M, Dietrich J, & McGonigle KM(2022). Comparison of a Chromogenic Urine Culture Plate System (UTid+) and Conventional Urine Culture for Canine and Feline Specimens. *Vetenary Science* 16,138-146. doi: 10.3390/vetsci9030138
- Chen Y, Cheng J, & Creamer S (2008). Inhibition of anaerobic digestion process: A review. *Bio*resource Technology 99, 4044 - 4064
- Donkin SS, Doane PH, & Cecava MJ (2013). Expanding the role of crop residues and biofuel co-products as ruminant feedstuffs. *Animal Front*iers 3, 5460.
- Ekka R, Vikas SV, & Kumar A (2016). Study of biogas production from poultry droppings waste.

 International Journal of Interdisciplinary Research Centre 2, 90 96.
- Garba B, & Uba A (2002). Biogas Generation from Ornamental Plants. Sokoto Energy Research Centre. Energy Commission of Nigeria Usmanu Danfodiyo University, Sokoto. *Nigerian Journal* of Renewable Energy 10, 61 - 62.
- Hagos K, Zong J, Li D, Liu C, & Lu X (2017). Anaerobic co-digestion process for biogas production: Progress, challenges and perspectives. *Renewable* and Sustainable Energy Reviews 76, 1485 - 1496.
- Haider U, Munir A, Ghafoor A, & Ali S (2020). design of biogas fermentation chamberand techniques to enrich bio-methanation, *Pak. J. Agri. Sci.*, 57, 1617-1627. doi:10.21162/PAKJAS/20.920http://www.pakjas.com.pk
- Hamilton DW (2022). Anaerobic digestion of manures:

 Methane potentials of waste materials. Oklahoma cooperative extension. Facts sheethttps://osufactOkstate.edu. Accessed on 02/09/2022
- Hawkes R, Pederson E, & Ngeleka M (2008). Mastitis caused by Bacillus anthracitic in a beef cow.

Canadian Vetinary Journal 49, 889 - 91.

- Iskalieva A, Yimmou BM, Gogate PR, Miklos HM, Horvath PG, & Csoka L (2012). Cavitation assisted
- delignification of wheat straw: A review. *Ultrasonics Sonochemistry* 19, 984 993.
- Jacob JH, Al-Fawwaz AT, & Al-Shira'h HH (2018). Evaluation and Optimization of Methane Production
- from Different Manure Types. *Jordan Journal of Biological Sciences*, 11, 323 327.
- Kartikey KG, Kamal RA, & Deepanshu R (2016). Current status of cow dung as a bioresource for
- sustainable development. *Bioresour. Bioprocess.* 3:28 -
- Kovács E, Wirth R, Maróti G, Bagi Z, & Rákhely G (2013). Biogas production from protein-rich biomass:
- Fed-Batch Anaerobic Fermentation of Casein and of Pig Blood and Associated Changes in Microbial
- Community Composition. *PLoS ONE* 8(10), e77265. doi:10.1371/journal.pone.0077265.
- Krause DO, Nagaraja TG, Wright AD, Callaway TR (2013). Board-invited review: Rumen microbiology:

Leading the way in microbial ecology. Journal of Animal Science 5 91, 331 341. doi:10.2527/jas.2012-5567.

Liang SJ, Sun J, Mahmood A, Basir A, Ashraf I, & Yang S (2021). Potential of Rapid Anaerobic

Fermentation on Animal Slurry for Biogas Production and Storage of Biogas. Polish Journal of

Environmental Studies 30, 247 - 256.

Lourenço A, Morgado F, Duarte LC, Roseiro LB, Fernandes MC, Pereira H, & Carvalheiro F (2021).

Delignification of Cistus ladanifer Biomass by Organosolv and Alkali Processes. Júnia Alves-

Ferreira. Energies 14. 1149. https://doi.org/10.3390/en1404112

Madigan M. Martinko J. Stahl D. & Clark D 2011. Brock Biology of Microorganisms. San Francisco, CA:

Pearson Education, Inc., Benjamin Cummings. pp734. McGninnis MR, & Borgers M(eds) (1980). Handbook of medical mycology. Academic Press, San Diego.

Moller HB, Sommer SG, & Ahring B (2004). Methane productivity of manure, straw and solid fractions of

manure. Biomass and Bioenergy 26, 485 - 95.

Mussoline W, Esposito G, Lens P, & Giordano A (2013). The anaerobic digestion of rice straw: A review.

Critical Review of Environmental Science & Technology 43, 895 - 915

Nizami AS, & Murphy JD (2010). What type of digester configurations should be employed to produce

biomethane from grass silage?. Renewable and Sustainable Energy Reviews, 14, 1558 - 1568.

Odonkor ST, & Mahami T (2020). Microbial air quality in neighbourhood near landfill sites: Implications

for public health. Journal of Environmental and Public Health. 1-10, doi.org/10.1155/2020/4609164

Panjaitan E. Sidauruk K, Manalu CJ, Saragih M, & Sianturi P (2022). Impact of agricultural waste

and NPK fertilizers on soil chemical properties, production and phosphorus uptake of sweet corn

plants on utisol soil. IOP conference series, Environmental Science, 10005012031, doi:10.1088/1755-1315/1/012030.

Pramasari DA, Sondari D, Rachmawati SA, Ningrum RS, & Sufiandi S (2021). The effect of alkaline-

autoclaving delignification on chemical component changes of sugarcane trash. Earth and

Environmental Science 759, 1-20. doi:10.1088/1755-1315/759/1/012010

Putri DA, Saputro R, & Budiyono R (2012). Biogas Production from Cow Manure. International Journal

of Renewable Energy Development, 1(2), 61-64.

Samani MS, Abdoli MA, Karbassi A, Pourzamani HR, & Rezaee M (2017). Effect of physical and chemical

operating parameters on anaerobic digestion of manure and biogas production, A review Journal of

Environmental Health & Sustainable Development 2,

Tapadia-Maheshwari S, Pore S, Engineer A, Shetty D, Dagar SS, & Dhakephalkar PK (2019). Illustration

of the microbial community selected by optimized process and nutritional parameters resulting in

enhanced biomethanation of rice straw without thermo-chemical pretreatment. Bioresour Technol.

289 13.

https://doi.org/10.1016/j.biortech.2018.09.107.

Varanasi P, Singh P, Auer M, Adams PD, Simmons BA, & Singh S (2013). Survey of renewable chemicals

-296.

produced from lingo cellulosic biomass during ionic liquid pretreatment. Biotechnology & Biofuels 6, 1 - 9.

Ye X, Chang Z, Qian Y, Pan J, Zhu J (2012). Investigation on large and medium scale biogas plants and

biological properties of digestate in jiangsu province. Nongye Gongcheng Xuebao/Transactions of

the Chinese Society of Agricultural Engineering, 28,

Yoshida K, Kametani K,& Shimizu N (2020). Adaptive identification of anaerobic digestion process for

biogas production management systems. Bioprocess Biosystem Engineering, 43,45-54

Zhang Y, Liu Z, Liu HT, Hui LF, Wang HM, & Liu HY (2019). Characterization of the liquefaction

residue from corn stalk and its biomass components using polyhydric alcohols with

phosphoric acid. Bio Resources, 14, 2684 - 2706. DOI: 10.15376/biores.14.2.2684-2706